



Topic : Early Flowering and High Yielding Mutants In
Ocimum sanctum Linn.

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Abstract

Early Flowering and High Yielding Mutants In *Ocimum Sanctum* Linn. were induced by physical (gamma rays) and chemical mutagens : SA (Sodium azide) and EMS (Ethylmethane sulphonate). The seeds were irradiated with 200,400,600 Gy gamma rays Seeds were also treated with SA (0.001%,0.002%,0.003%) and EMS (0.1%,0.2%,0.4). Population was screened carefully in M2 and M3 generations to isolate the mutants with early flowering and reduced maturation period and high yielding. Early flowering mutants were recorded in the M2 and M3 generation. The physical mutagen (gamma rays), as well as both chemical mutagens (SA and EMS) were found to induce high yielding mutants.

Introduction

The plants in the mutagen treated populations which commenced flowering about 15-20 days earlier than control plants were categorized as Early flowering. In the control plants, flowering commenced on 40-50 days after germination, whereas in the early flowering mutants, the flowering was found to start on 19th to 29th day after germination. The physical mutagen (gamma rays) as well as both chemical mutagens (SA and EMS) were found to induce high yielding mutants. These high yielding mutants were characterized by having considerably high seed yield, more number of branches and some of these were also taller than the control plants.

Keywords :

Physical and chemical mutagens Early flowering mutants, High yielding mutants, *Ocimum sanctum*.

Material and Methods

Dry seeds of *Ocimum sanctum* Linn. of uniform size and shape were treated in gamma cell with 60 CO as gamma ray source @ 100 Gy per minute at the Department of chemistry, RTM Nagpur University Campus, Nagpur. They were irradiated with 200, 400, 600 and 800 Gy doses of gamma rays. Seeds without irradiation served as control. Two chemical mutagens, sodium azide (E.Merck, Germany) and ethylmethane sulphonate (Sigma, USA) were used for inducing mutations. Seeds were presoaked for

5 and 10 hours in distilled water, surface dried with blotting paper and subjected to chemical mutagens.

The dry as well as pre. soaked seeds were treated with 20 ml aqueous solutions of 0.0, 0.001,0.002 and 0.003% Sodium azide (SA) and 0.00,0.1,0.2 and 0.4 % ethyl methane sulphonate (EMS) for 18 hours with uniform and continuous shaking on orbital shaker at 22 10C in 50 ml flask. All other conditions were similar. Treated seeds were thoroughly washed in running tap water, soaked in 50 ml of glass distilled water for 2 hours, again washed in running tap water and surface dried.

One hundred seeds of each treatment were sown in pots and the seedlings were transplanted in the field at sixth leaf stage. M2 and M3 populations were screened for early flowering and high yielding mutants.

Results and Discussion

The plants in the mutagen treated populations which commenced flowering about 15-20 days earlier than control pants were categorized as 'Early flowering'. In the control plants, flowering commenced on 40-50 days after germination, whereas in the early flowering mutants, the flowering was found to start on 19th to 29th day after germination. In some of the early flowering mutants, the number of branches and the weight of seeds were found to be more than control. The frequency of these mutants ranges from 0.28 to 0.27% in M2 and M3 0.38 to 0.26 in M3 in gamma ray treated population. In SA treated population, the frequency was 0.66 (Dry set 0.001%), 1.33 (5hpsw 0.001%) and 0.37% (10 hpsw 0.002%) in M2 generation and in M3 generation, it ranges from 0.8 to 0.49% (Dry set), 0.86 to 0.25% (5hpsw set), and 0.26% (10 hpsw 0.002%) respectively. In EMS treated population, the frequency was 0.19% (Dry set 0.1%), 0.16% (5 hpsw 0.4%) in M2 generation and in M3 generation, it ranges from 0.55 to 0.30 (Dry set), 0.53 to 0.8% (5hpsw set) and 0.19 to 0.31 (10hpsw set) respectively. All the three mutagens were observed to induce early flowering mutants with diferent frequencies. Early flowering mutants were also recorded in the M3 generation. (Table-1)

The physical mutagen (gamma rays), as well as both chemical mutagens (SA and EMS) were found to induce high yielding mutants. These high yielding mutants were characterized by having considerably high seed yield, more number of branches and some of these were also taller than the control plants. The plants height of these plants ranged between 66.8 to 103.1 cm and the number of branches varied from 18 to 38. The seed weight of per plants was 0.798 - 0.982 g. The frequency of these mutants was 0.28% in the gamma ray treated M2 population, and ranged between 0.74 to 0.33% in SA treatment. In EMS treated M2 population frequency was 0.19%. In the M3 generation also, the mutants were noted. (Table - 1)

Discussion

Earliness of flowering is one of the desirable characters, which can be reliably obtained in mutation experiments. In *Ocimum sanctum*, number of early flowering mutants have been obtained with the use of the gamma rays, SA and EMS. Early flowering mutants have been reported in barley by Hagberg (1969), Domini and Devraux (1970), Scholz (1971), Bansal (1971,1972), Ibrahim and Sharan (1974), Yameguchi et al (1974), Stephanov and Gorastev (1976), Hussein et al (1979, 1980) and Ukai and Yamashita (1980). Early flowering mutants in different plants were also reported. Marie (1970) with X- rays, gamma rays and EMS, Tanaka (1969) with gamma rays, Miah et al (1970) with gamma rays. Haq et al (1971) with gamma rays. Reddy and Reddy (1971, 1972) with gamma rays, neutrons and dES Ram (1974) with gamma rays, Ismachin and Mikaelson (1976) with gamma rays, Kaul (1978) with gamma rays and Afsar et al. (1980) with SA very early and early flowering mutants have been reported by Kumar and Dubey (1998) in *Lathyrus* with gamma rays, individually, as well as, in combination with EMS or dES. George and Nayar (1973), in linseeds, have attributed earliness in flowering to the physiological changes caused by irradiation.












High yielding mutants were also obtained by physical and chemical mutagen treated population in *Ocimum sanctum*. Aastveit (1966) and Pollhamer (1966, 1967) reported high yielding mutants in *Hordeum vulgare*. Similar reports were made by Tanaka (1968), Saini and Sharma (1970), Pawar (1971), Kaul (1978), Ismail and Ahmad (1979) in *Oryza sativa* Jech (1966), Khvostova (1967), Mar'yushkin et al (1977), Siddiqui and Arain (1974), Sawhney and Sharma (1979), Sichkar et al (1980) and Siddiqui et al (1981) also reported high yielding mutants

in *Triticum aestivum*. High yielding mutants were also reported by Kanaklata (1995) in *Cyamopsis tetragonoloba*, Landge (2000) in *Brassica napus* and Girhe and Choudhary (2002) in *Lathyrus sativus*.

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